

KINETIC STUDIES OF TYROSINE KINASE INHIBITION BY ERBSTATIN

Sir:

Erbstatin was isolated from the culture filtrate of *Streptomyces* sp. MH435-hF3 as a potent inhibitor of epidermal growth factor (EGF) receptor associated tyrosine protein kinase¹. It inhibits EGF receptor-autophosphorylation with IC_{50} of 0.55 $\mu\text{g/ml}$ *in vitro*, but does not inhibit either cyclic AMP dependent protein kinase¹ or protein kinase C (Dr. H. HIDAOKA, personal communication). It was also shown to inhibit EGF-stimulated receptor-autophosphorylation in cultured A431 cells and the autophosphorylation of p60^{src} in Rous sarcoma virus-infected rat kidney cells². Besides autophosphorylation of the above proteins, the synthetic peptide (Arg-Arg-Leu-Ile-Glu-Asp-Ala-Glu-Tyr-Ala-Ala-Arg-Gly (RR-SRC)) was found to be a good substrate for EGF receptor associated tyrosine kinase³, therefore, using this peptide as a substrate we have studied the mechanism of enzyme inhibition by erbstatin.

EGF receptor associated tyrosine kinase was partially purified from human epidermoid carcinoma A431 cells⁴. A431 cells were grown in DULBECCO's modified EAGLE's medium supplemented with 5% calf serum, washed with cold DULBECCO's phosphate buffered saline, scraped off with rubber policeman, and centrifuged. Then, the cells were homogenized in 20 mM *N*-2-hydroxyethylpiperazine-*N'*-2-ethanesulfonic acid, purchased from Sigma (HEPES) buffer (pH 7.4) containing 1 mM MgCl_2 and 5 mM KCl. The homogenate was centrifuged at 15,000 $\times g$ for 5 minutes, and the supernatant was re-centrifuged at 100,000 $\times g$ for 30 minutes. The pellet obtained as a membrane fraction was solubilized for 60 minutes at 4°C in 25 mM HEPES buffer (pH 7.4) containing Triton X-100 1%, glycerol 10%, leupeptin 0.15 mg/ml, aprotinin 0.15 mg/ml and phenylmethylsulfonyl fluoride (PMSF) 0.2 mM. Insoluble material was removed by centrifugation at 100,000 $\times g$ for 60 minutes. The supernatant was then diluted so that the Triton X-100 concentration was 0.05% and passed four times through a column containing 2 ml of wheat germ agglutinin-agarose (Seikagaku Kogyo Co., Ltd.) at 2°C. The column was extensively washed with the Triton X-100

solution containing Triton X-100 0.05%, glycerol 10%, PMSF 0.2 mM, NaCl 0.5 M and HEPES buffer 40 mM (pH 7.4). The EGF receptor was eluted with the buffer above added with 0.3 M *N*-acetylglucosamine, and dialyzed against the Triton X-100 solution. Tyrosine kinase reactions were carried out in a final volume of 60 μl containing HEPES-NaOH 20 mM, pH 7.4, MnCl_2 1 mM, EGF 100 ng, the purified EGF receptor (12 μg protein) and indicated concentrations of inhibitors, RR-SRC (Peninsula Laboratory) and [γ -³²P]ATP. The kinase activities were measured by a slight modification of the method described by GLASS *et al.*⁵. The EGF receptor was first incubated with EGF at 23°C for 30 minutes before assay of kinase activity. The kinase reactions were initiated by the addition of RR-SRC and [γ -³²P]ATP, and the reaction mixture was incubated for 10 minutes at 23°C. The reactions were terminated by addition of 25 μl of 10% TCA and 6 μl of bovine serum albumin (10 mg/ml). Precipitated proteins were removed by centrifugation and 40 μl aliquot of the supernatant was spotted on a Whatman P81 paper (2 \times 2 cm) which were immediately immersed in 30% AcOH at 25°C. The papers were washed four times for 15 minutes in 15% AcOH, for 5 minutes in Me_2CO , and dried. The radioactivity was counted with a liquid scintillation counter.

As shown in Fig. 1(A), the inhibitory pattern in the Lineweaver-Burk plot of erbstatin vs. peptide was a typical competitive inhibition, in which each line met together on the *y*-axis. The inhibitory pattern of erbstatin vs. ATP was noncompetitive as shown in Fig. 1(B), in which each line met on the *x*-axis. From the Dixon plot analysis the K_i value was found to be about 5.58 μM . (*E*)-2-(2,3-dihydroxyphenyl)vinylformamide, a structural analogue of erbstatin, gave the same pattern of kinetics as erbstatin in tyrosine protein kinase inhibition with K_i value of 2.23 μM (data not shown). Orobol was first isolated from the *Streptomyces* culture broth as an inhibitor of dopa decarboxylase⁶ and then isolated again in the course of screening for tyrosine protein kinase inhibitor¹⁷. It has an isoflavonoid structure and inhibits EGF receptor tyrosine protein kinase with IC_{50} of 3.0 $\mu\text{g/ml}$ *in vitro*. As shown in Fig. 2(A) inhibition of tyrosine protein kinase by orobol was competitive to ATP. The Lineweaver-Burk plot of orobol vs. the peptide showed neither noncom-

Fig. 1.

(A) A Lineweaver-Burk plot of enzyme kinetics showing competitive inhibition by erbstatin against peptide substrate. Inhibitor concentrations are 0 (\circ), 11.16 (\bullet), 22.3 (Δ) and 33.5 (\square) μM , respectively.

(B) A Lineweaver-Burk plot of enzyme kinetics showing noncompetitive inhibition by erbstatin against ATP. Inhibitor concentrations are same as (A).

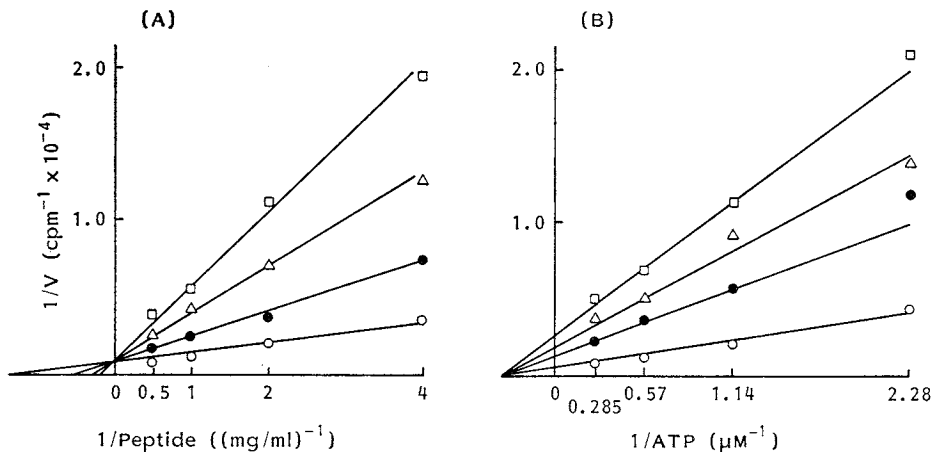
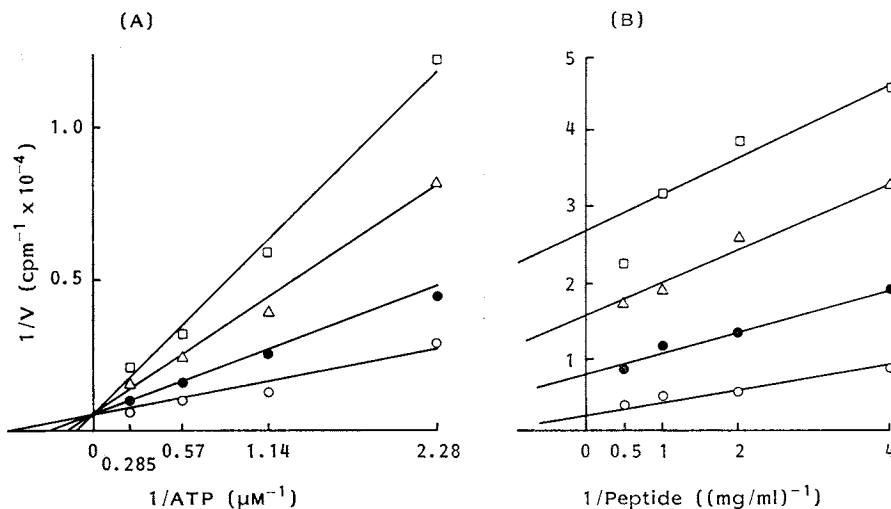


Fig. 2.

(A) A Lineweaver-Burk plot showing competitive inhibition by orobol against ATP. Inhibitor concentrations are 0 (\circ), 1.75 (\bullet), 3.49 (Δ) and 5.24 (\square) μM , respectively.

(B) A Lineweaver-Burk plot showing not competitive inhibition by orobol against peptide substrate. Inhibitor concentrations are same as (A).



petitive nor uncompetitive as shown in Fig. 2(B).

Thus, erbstatin competes with the peptide substrate, and the mechanism of inhibition is clearly different from that of orobol which competes with ATP. The EGF receptor associated tyrosine kinase was reported to act in the Ordered Bi Bi mechanism, in which the peptide came first and ATP second to the enzyme active site, since

the dipeptide Tyr-Arg, a weak tyrosine protein kinase inhibitor (K_i 600 μM) competed with the synthetic peptide and noncompeted against ATP, and since ADP, a product of the kinase reaction, was a linear noncompetitive inhibitor with respect to ATP and a linear competitive inhibitor with respect to peptide⁷. Our result confirmed this mechanism using the more potent inhibitor

erbstatin, and also erbstatin was confirmed to attack the enzyme directly.

The structure of erbstatin apparently resembles tyrosine, and it is possible that erbstatin resembles to a greater extent the specific conformation of tyrosine within the peptide. Genistein^{8,9)} and amiloride¹⁰⁾ which inhibit tyrosine kinase and H-8¹¹⁾ and K-252 compounds¹²⁾ which inhibit protein kinase C are all competitive to ATP. Erbstatin is a unique tyrosine protein kinase inhibitor competing with the peptide substrate.

MASAYA IMOTO
KAZUO UMEZAWA
KUNIO ISSHIKI
SETSUKO KUNIMOTO
TSUTOMU SAWA
TOMIO TAKEUCHI
HAMAO UMEZAWA

Institute of Microbial Chemistry,
3-14-23 Kamiosaki, Shinagawa-ku,
Tokyo 141, Japan

(Received May 22, 1987)

References

- 1) UMEZAWA, H.; M. IMOTO, T. SAWA, K. ISSHIKI, N. MATSUDA, T. UCHIDA, H. INUMA, M. HAMADA & T. TAKEUCHI: Studies on a new epidermal growth factor-receptor kinase inhibitor, erbstatin, produced by MH435-hF3. *J. Antibiotics* 39: 170~173, 1986
- 2) IMOTO, M.; K. UMEZAWA, T. SAWA, T. TAKEUCHI & H. UMEZAWA: *In situ* inhibition of tyrosine protein kinase by erbstatin. *Biochem. Int.* in press
- 3) CASNELLIE, J. E.; M. L. HARRISON, L. J. PIKE, K. E. HELLSTRON & E. G. KREBS: Phosphorylation of synthetic peptides by a tyrosine protein kinase from the particulate fraction of a lymphoma cell line. *Proc. Natl. Acad. Sci. U.S.A.* 79: 282~286, 1982
- 4) AKIYAMA, T.; T. KADOOKA & H. OGAWARA: Purification of the epidermal growth factor receptor by tyrosine-sepharose affinity chromatography. *Biochem. Biophys. Res. Commun.* 131: 442~448, 1985
- 5) GLASS, D. B.; R. A. MASARACCHIA, J. R. FERAMISCO & B. E. KEMP: Isolation of phosphorylated peptides and proteins on ion exchange papers. *Anal. Biochem.* 87: 566~575, 1978
- 6) UMEZAWA, H.; H. TOBE, N. SHIBAMOTO, F. NAKAMURA, K. NAKAMURA, M. MATSUZAKI & T. TAKEUCHI: Isolation of isoflavones inhibiting dopa decarboxylase from fungi and streptomycetes. *J. Antibiotics* 28: 947~952, 1975
- 7) ERNEUX, C.; S. COHEN & D. L. GARBERS: The kinetics of tyrosine phosphorylation by the purified epidermal growth factor receptor kinase of A431 cells. *J. Biol. Chem.* 258: 4137~4142, 1983
- 8) OGAWARA, H.; T. AKIYAMA, J. ISHIDA, S. WATANABE & K. SUZUKI: A specific inhibitor for tyrosine protein kinase from *Pseudomonas*. *J. Antibiotics* 39: 606~608, 1986
- 9) OGAWARA, H.; T. AKIYAMA, J. ISHIDA, S. WATANABE & K. SUZUKI: A specific inhibitor for tyrosine protein kinase. 14th Int. Cancer Congr. Abstracts of Lectures, Symposia and Communications. Vol. 3, p. 1139, Budapest, Aug. 21~27, 1986
- 10) DAVIS, R. J. & M. P. CZECH: Amiloride directly inhibits growth factor receptor tyrosine kinase activity. *J. Biol. Chem.* 260: 2543~2551, 1985
- 11) HIDAKA, H.; M. INAGAKI, S. KAWAMOTO & Y. SASAKI: Isoquinolinesulfonamides, novel and potent inhibitors of cyclic nucleotide dependent protein kinase and protein kinase C. *Biochemistry* 23: 5036~5041, 1984
- 12) KASE, H.; K. IWASHASHI, S. NAKANISHI, Y. MATSUDA, K. YAMADA, M. TAKAHASHI, C. MURAKATA, A. SATO & M. KANEKO: K-252 compounds, novel and potent inhibitors of protein kinase C and cyclic nucleotide-dependent protein kinases. *Biochem. Biophys. Res. Commun.* 135: 397~402, 1986